



THINK AGAIN

Insights & Perspectives

Asymmetrical sex reversal: Does the type of heterogamety predict propensity for sex reversal?

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Abstract

Sex reversal, a mismatch between phenotypic and genetic sex, can be induced by chemical and thermal insults in ectotherms. Therefore, climate change and environmental pollution may increase sex-reversal frequency in wild populations, with wide-ranging implications for sex ratios, population dynamics, and the evolution of sex determination. We propose that reconsidering the half-century old theory “Witschi’s rule” should facilitate understanding the differences between species in sex-reversal propensity and thereby predicting their vulnerability to anthropogenic environmental change. The idea is that sex reversal should be asymmetrical: more likely to occur in the homogametic sex, assuming that sex-reversed heterogametic individuals would produce new genotypes with reduced fitness. A review of the existing evidence shows that while sex reversal can be induced in both homogametic and heterogametic individuals, the latter seem to require stronger stimuli in several cases. We provide guidelines for future studies on sex reversal to facilitate data comparability and reliability.

KEYWORDS

ectotherm sex determination, endocrine-disruptor pollution, feminization, masculinization, sex-chromosome evolution, sex-reversing temperature

INTRODUCTION

Ectothermic vertebrates feature a variety of sex-determination systems including both genetic and environmental types.^[1–3] The most widespread variants of genetic sex determination are male-heterogametic (XX/XY) and female-heterogametic (ZW/ZZ) systems, displaying a variety of, often homomorphic, sex chromosomes. Across ectotherm vertebrates, certain environmental conditions, including temperature and the presence of several chemical compounds, can

favor the development of one phenotypic sex, even in species possessing genetic sex determination. This leads to sex reversal, producing genetic males with female phenotype or vice versa. Sex reversal has been documented in wild populations of all major ectothermic vertebrate taxa.^[2,4–8] Theoretical models predict that global climate change and environmental pollution is expected to increase sex-reversal frequency in these taxa, potentially affecting sex-chromosome evolution, causing skewed sex ratios and even leading to extinction.^[1,9–12] Therefore, understanding mechanisms behind the evolution of sex reversal is an important and urgent challenge. Theoretical works have shown that male- and female-heterogametic systems might respond differently to

Abbreviations: DHT, dihydrotestosterone; E2, 17 β -estradiol; EE2, 17 α -ethynylestradiol.

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environmental changes via sex reversals,^[10,11,13] but there is a significant knowledge gap yet to be filled with empirical data.

About 60 years ago, based on experiments applying exogenous sex hormones to a few amphibian species, Witschi and colleagues^[14] recognized that it was predominantly the homogametic sex (XX or ZZ) that was susceptible to sex reversal – a concept sometimes referred to as Witschi's rule (Box 1). Roughly 20 years later, Adkins-Regan^[15] came to a similar conclusion based on reviewing data from fish, amphibians, reptiles, and further taxa. However, laboratory experiments across ectothermic vertebrates successfully produced both sex-reversed XY females and ZW males, and these were even fertile in some species.^[16–20] Therefore, some authors see Witschi's rule as disproved,^[21,22] while others maintain that sex reversal is restricted to the homogametic sex, acknowledging that there are counterexamples with no explanation.^[2,13] Clarifying this issue empirically would be important for understanding which species are susceptible to sex reversal induced by specific environmental stimuli: Witschi's rule predicts higher vulnerability to male-to-female sex-reversing effects such as xenoestrogens in ZW/ZZ compared to XX/XY systems. By contrast, XX/XY systems should be more inclined to female-to-male sex reversal. Furthermore, whether the heterogametic sex is resistant to sex reversal or not is important also for our theoretical understanding of the evolution of sex-determination systems: some models dealing with transitions between these systems assumed that only certain sex-chromosome genotypes can undergo sex reversal,^[11,13] while others made no such assumption.^[10,23–27]

After knowledge kept gathering on the evolution of sex chromosomes and sex-determination systems during the past century, it was suggested that natural selection could cause the pattern described by Witschi's rule.^[13] The key seems to be that if sex-reversed individuals participate in breeding, new combinations of the sex chromosomes (YY or WW) can emerge in their progeny (Box 2). These new genotypes may possess reduced fitness due to degeneration of the genetic content of the hemizygous chromosome,^[13,25] driven by accumulation of deleterious mutations^[28–30] and sex-antagonistic genes^[31,32] (but see^[33] and^[34]). These new genotypes can only be produced if the sex-reversed parent is heterogametic (XY female mating with XY male, or ZW male mating with ZW female). Thus, reduced fitness of the new genotype may lead to selection against sex reversal in the heterogametic sex (Box 2). As a result, the homogametic sex (XX or ZZ) may be more susceptible to sex reversal compared to the heterogametic sex. This potential evolutionary mechanism has been mentioned in several reviews on environmental sex reversal^[2,35–37]; however, similarly to earlier attempts,^[15,21,38] none of these articles offered a robust explanation why some species conformed to Witschi's rule while others did not.

We propose that the apparent contradiction between Witschi's rule and empirical findings may be resolved by acknowledging that the propensity for sex reversal may vary on a gradual scale (Figure 1A) and may be shaped by various factors. In ectotherm vertebrates, different sex-determination systems dynamically replace each other as species evolve,^[39] and sex reversal of homogametic individuals may be an important driver of these transitions.^[13] Degeneration

Box 1: Witschi's rule

More than half a century ago, reviewing experiments that applied various sex-reversing treatments to amphibians, Emil Witschi and colleagues^[14] came to the conclusion that species can be classified into one of two reaction types: "In the first type, androgens produce complete and durable masculinization of genetically female embryos . . ." "In the second reaction type, . . . androgens produce either no major effects or, in some instances, they feminize genic males." Based on the knowledge available at the time, the authors assumed that the first reaction type was characteristic of anurans with male-heterogametic (XX/XY) sex-determination (based on ranid and hyloid frogs), while the second type was represented by "several families of primitive anurans (discoglossids, xenopins) and all urodeles," that were all assumed to possess female-heterogametic (ZW/ZZ) sex determination. With other words, sex-reversal could be induced in homogametic (XX or ZZ) individuals, while the heterogametic sex (XY or ZW) appeared resistant. They published their observation in a short article and did not provide detailed explanation on this pattern. One year later, Witschi published a review^[38] where he concluded that this pattern probably reflected genealogy across vertebrates, with noticeable exceptions.

Today we know that not all urodeles are female heterogametic, and not only "primitive anurans" possess this type of sex determination.^[46,48] In fact, sex-determination systems have been replacing each other during the evolution of amphibians, similarly to fishes and reptiles.^[39,48,86] As seemingly controversial information kept gathering on the inducibility of sex reversal across ectotherms,^[15–21,56] Witschi's rule became largely forgotten by the scientific community. Theoretical and empirical works from the past decade suggested that deleterious mutations present on the Y and W chromosomes may restrict sex reversal to the homogametic sex in reptiles^[2,13] (ultimately causing the same pattern that was observed by Witschi and colleagues in amphibians). Still, the various counterexamples that were reported in both taxa remained unexplained to date.

of the Y or W chromosome, and thus the strength of selection for restricting sex reversal to the homogametic sex, should gradually increase with the evolutionary age of the sex chromosomes (Box 2). Therefore, heterogamety-based differences might be less prominent in younger sex-determination systems. Furthermore, resilience to certain external factors may have physiological limits, and consequently, increased exposure to these factors might lead to sex reversal despite the system's relative resistance to it (Figure 1A). Comparison of sex-determination systems is further complicated by the possibility of phylogenetic inertia in sex-reversal sensitivity to

Box 2: Evolution of sex determination via sex reversal

At least 19 turnovers of male- and female-heterogametic sex-determination systems occurred during amphibian evolution,^[46] and sex determination mechanisms are even more variable in fish and reptiles.^[39,48,86] These turnovers may or may not be accompanied by the replacement of old sex chromosomes with new (formerly autosomal) ones, while many sex-chromosome turnovers remain hidden from the researchers' eyes behind the same type of heterogamety.^[92] There are numerous theoretical works concentrating on potential causes and genetic mechanisms of turnovers between sex-determination systems,^[25,31,93] and some of these indicate that environment-induced sex reversal might play an important role.^[10,13,27,36,94] It is a generally accepted rule that after recombination stops between the different sex chromosomes, the chromosome which never occurs in homozygotic form (Y in XX/XY systems and W in ZW/ZZ systems) will not be purged from deleterious mutations, which are accumulating over evolutionary time until a theoretical point where the chromosome cannot fulfill its purpose anymore.^[29,31,32] However, recombination appears to occur more frequently in females in various species, and according to the "fountain of youth" hypothesis, sex reversal may provide an opportunity for the Y chromosome to recombine with the X in sex-reversed XY females, hence slowing its degeneration process.^[95] Another important outcome of sex reversal by heterogametic individuals is the production of offspring with new genotypes: if a sex-reversed XY female mates with a sex-concordant XY male, 25% of their offspring will have YY genotype. Similarly, 25% WW offspring will be produced by a sex-reversed ZW male and a sex-concordant ZW female. Depending on the progress of Y or W degeneration, offspring with the new genotype might experience no detectable disadvantage, or may suffer negative fitness consequences on various levels, such as reduced fertility, lower survival rate or complete lethality.^[16,21,96] In these latter cases, selection is expected to favor those XY and ZW individuals that are resistant to sex reversal, and therefore, retain the phenotypic sex which is encoded by their sex chromosomes under sex-reversing conditions experienced by the population. By contrast, sex reversal in homogametic individuals (XX or ZZ) may be neutral or even advantageous: for example, sex-reversed ZZ females may lay more eggs than ZW females do,^[40] and XX males may produce female-biased progeny when the population sex ratio is male-biased due to climate warming.^[10] The propensity to develop as phenotypic male or female is underlay by heritable genetic,^[3,8,97-99] and epigenetic variation,^[8,100] providing diverse grounds for selection of sex-reversal resistance in natural systems.^[43] In line with this, sex-reversal propensity was found to be heritable in multiple species.^[37,40] Consequently, sex-reversal propensity may be reduced to various extents in the heterogametic sex across contemporary species and populations, which we may detect as "asymmetrical sex reversal".

conclusive comparisons of sex-reversal propensity between XX/XY and ZW/ZZ systems may be made by controlled experiments that manipulate environmental conditions during sex determination.

Experimental evidence for "asymmetrical sex reversal"

Amphibians, the taxon in which Witschi has originally discovered his rule, offers an ideal group for testing "asymmetrical sex reversal." Genetic sex determination underlies sexual development in all amphibian species studied so far, unlike in fishes and reptiles where many species seem to have temperature-dependent sex determination with little genetic influence.^[46-48] We searched the literature for sex-reversal and phenotypic sex-ratio data from laboratory experiments that were carried out on anuran amphibian species with either XX/XY or ZW/ZZ sex-determination system, focusing on the sex-reversing effects that were most often studied in this regard: developmental temperature, sex hormones, and anthropogenic chemicals with endocrine-disrupting effects. From the latter group, we chose the two compounds that have been studied most frequently: the contraceptive 17 α -ethynylestradiol (EE2) and the herbicide atrazine.^[12] Detailed searching methods are described in Supplement 1, and the data extracted

in Supplementary Table 1. We found only four experiments in which anuran species with both male and female heterogamety were studied for sex-reversal propensity,^[22,49-51] although heterogamety was not in their focus. Other studies were usually restricted to a single species. Experimental methods differed greatly across studies, including the applied concentrations of the same compounds as well as water temperature. Because genetic sex markers have been established for only a handful of amphibian species so far (e.g.,^[4-7,41]), in the vast majority of studies sex reversal was inferred based on biased phenotypic sex ratios produced by specific treatments. Several relevant experimental conditions, such as treatment duration or mortality rates, were unclear in numerous instances, especially among broadly cited publications from the previous century.^[52-54] For these reasons, we judged that formal meta-analyses would be unfeasible with the currently available, highly heterogeneous data.

The only treatment type where sex reversal was confirmed by genetic sex markers in both male- and female-heterogametic anurans was the administration of EE2 in the rearing water. Such studies have been carried out in two species with ZW/ZZ, and three species with XX/XY system (Figure 1B). The reported sex-reversal frequencies are in agreement with the theory of "asymmetrical sex reversal": genetic males became phenotypic females in ZW/ZZ species at lower EE2 concentrations compared to XX/XY species. In the anuran *Glandirana*

rugosa, different populations feature different sex-determination systems.^[55] Using a variety of sex hormones, sex reversal could be induced only in ZZ individuals in the ZW/ZZ populations, while it was absent in the population with heteromorphic X and Y chromosomes, and it occurred in both XX and XY individuals in populations with homomorphic sex chromosomes.^[55] All these data on proven cases of sex reversals support that the homogametic sex has higher propensity to undergo sex reversal, especially when the two sex chromosomes are more diverged. Although a study on reptiles^[1] concluded that sex-chromosome heteromorphy does not constrain the sensitivity to sex reversal, this conclusion was based on the finding that high doses of 17 β -estradiol (E2) injected into the eggs caused 100% female phenotype in two turtle species with XX/XY sex determination, regardless of their sex chromosomes being heteromorphic or homomorphic. Low sample sizes and the lack of genetic sexing both limit the interpretation of these results.

Lacking data on genetically confirmed sex reversals in most species, tentative speculations can be made based on phenotypic sex ratios (Figure 2 and 3). Out of 18 anuran species for which we found sex-ratio data from water-temperature, sex-hormone, or atrazine treatments, only four featured ZW/ZZ sex-determination system. Treatments with testosterone, dihydrotestosterone (DHT) and high temperature caused sex-ratio bias toward males, but complete or near-complete elimination of phenotypic females (\geq 98% males) at higher treatment values was achieved only in XX/XY species (Figure 2). Treatments with E2 and atrazine tended to cause sex-ratio bias toward females, and ZW/ZZ species produced the strongest responses: only ZW/ZZ species reached 100% female sex ratios for E2 (excepting a single XX/XY species, *Pseudacris triseriata*) and high female bias (< 30% males) for atrazine (Figure 3). Majority of the studies we overviewed accounted for the presence of intersex individuals, although the definition of intersexuality differed between articles: in general, it included individuals with one ovary and one testis, or gonads with mixed-sex tissue based on either gross morphology or histology. The proportion of intersex individuals can vary greatly between and within species (Figure 1B, 2, and 3), sometimes even exceeding 50% of the treated individuals; many of these cases are likely signs of incomplete sex reversal and might indicate limited sex-reversal ability in the genetic sex affected by the applied treatment. However, intersex and sex reversal may also occur independently of each other,^[5,57] as intersex may be a natural phase of gonad development in some species of amphibians as well as fish^[12,58] and reptiles.^[59]

Sex-ratio data suggest that some species might be less susceptible to sex reversal compared to others with the same sex-determination system. For example, while phenotypic sex ratio in two other XX/XY species (*Pseudacris triseriata* and *Hyperolius viridiflavus*) was strongly affected by exogenous testosterone treatment, it was not distorted in *Hyla arborea*, the third such species, by 100 000 ng/l, twice the concentration that already caused 100% male phenotype in *Pseudacris triseriata* (see Figure 2B). However, this outlying lack of sex-ratio bias in *Hyla arborea* might be a treatment-specific outcome, since testosterone can be modified into estrogen by α -aromatase in the endocrine system; and testosterone treatments may even cause “paradoxical” male-to-female

sex reversal in some species, while nonaromatizable androgens, such as DHT, cause female-to-male sex reversal in them.^[60,61] Similar heterogeneity was found for fishes where methodological differences across studies accounted for much more interspecific variation in sex-reversal inducibility than biological differences.^[44] Furthermore, even within-species differences can occur in apparent sex-reversal inducibility: for the best-studied species, *Xenopus laevis*, sex ratios observed after similar treatments greatly differed between studies (e.g.,^[62] vs.^[63]). Such differences may stem from discrepancies in the experimental set-up, sample size, or other methodological details (see below). Thus, while the patterns in Figures 1B, 2, and 3 are largely in agreement with the idea that the homogametic sex is more susceptible to sex reversal, there is also noise in these patterns, and understanding the sources of this variation would be important for understanding what makes certain animals more susceptible to sex reversal than others.

The devil in the details: How to choose suitable methods?

In order to enable systematic comparison of the responsiveness of different sex-determination systems to sex-reversing effects, future studies should apply the same experimental design in both XX/XY and ZW/ZZ species concurrently. This will minimize the risk that differences between species are confounded by uncontrolled differences in the circumstances (such as varying relationships between nominal and actual treatment concentrations; Supplementary Table 1). Ideally, such experiments should include multiple treatments within the range of ecologically relevant concentrations or temperatures, to facilitate the recognition of ranges where sex-reversal inducibility differs between the two sex-determination systems (see Figure 1). Pairwise comparison of closely related XX/XY and ZW/ZZ species (or populations of the same species^[55]) would be best fitting for this purpose; see Figure 4 for a nonexhaustive collection of such candidate species across ectotherm vertebrates. Once we have enough data from such experiments, quantitative meta-analyses of the within-experiment differences will be executable to ascertain whether the type of heterogamety is a consistent determinant of sex-reversal propensity.

Even when it is not possible to include more than one species in an experiment on sex reversal, there is much researchers can do to make future findings more directly comparable among each other and clearer to interpret. We should endeavor to identify sex reversal correctly. When the conclusions are drawn solely from phenotypic sex ratios, it should be born in mind that such conclusions can be strongly affected by sex-biased mortality^[57,64] and stochasticity stemming from low sample sizes. Therefore, mortalities and sample sizes should always be clearly reported. Preferably, sex-reversed individuals should be identified by genetic sexing,^[49,57,64] and for this, development of genetic sex markers for those many thousands of species where such markers are not yet available is an inevitable challenge.

When designing sex-reversal experiments and reporting the data, several methodological aspects should be considered explicitly. Different species can have very different pace of ontogeny and the sensitive

Sex determination: — XX/XY — ZW/ZZ Proportion of intersex (♂): ● 0 ● .1 ● .2 ● .3 ● .4 ● .5 ● .6 ● Present ○ Unknown

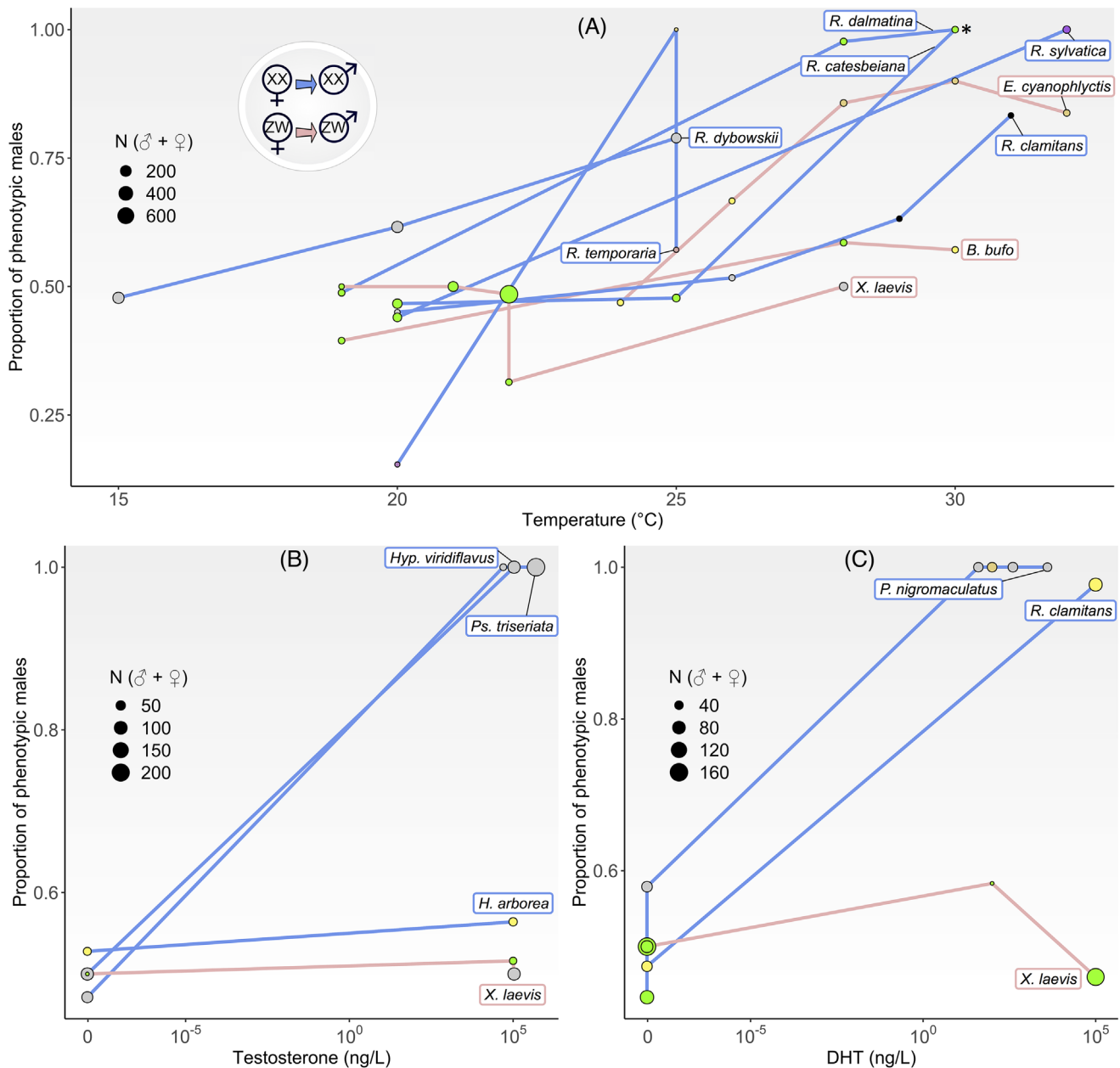


FIGURE 2 Phenotypic sex ratios (i.e., proportion of males among individuals with unambiguous sexual phenotype) reported from anurans exposed to different temperatures (A), or different concentrations of testosterone (B), or DHT (C) as tadpoles. Within each panel, dot sizes are proportional to the number of animals with unambiguous sexual phenotype; dot colors indicate the proportion of intersex individuals among all animals examined for intersexuality. Supposed direction of sex reversal across all panels is denoted in the large white circle in Panel (A), where the symbols ♂ and ♀ stand for male and female phenotype, respectively. Displayed anuran genera are: *Bufo* (B.), *Euphlyctis* (E.), *Hyla* (H.), *Hyperolius* (Hyp.), *Pelophylax* (P.), *Pseudacris* (Ps.), *Rana* (R.) and *Xenopus* (X.). Two overlapping data points are marked by an asterisk: the proportion of intersex individuals at 30°C was 0 in *Rana dalmatina* and 0.56 in *Rana catesbeiana* (A). For data and references see Supplementary Table 1

period to sex-reversing effects can also vary between them.^[22,55,57,65] Therefore, treatment periods should include the time frame when sex reversal may be induced in each species to be compared. To ensure this, we need data on the timing of the sensitive window of each species. When such information is lacking, exposure to the sex-reversing treatment should either last for a long period during embryonic/larval development or applied in several different, shorter periods. How-

ever, we should also keep in mind that shorter treatments may be more environmentally relevant when applying some sex-reversing stimuli such as heat waves,^[66,67] while longer treatments may better simulate natural conditions with others such as persistent chemical pollutants. Another issue of timing is the diagnosis of phenotypic sex. In sex-reversal experiments, phenotypic sex is usually identified based on gonad morphology of dissected young animals, for example, at or

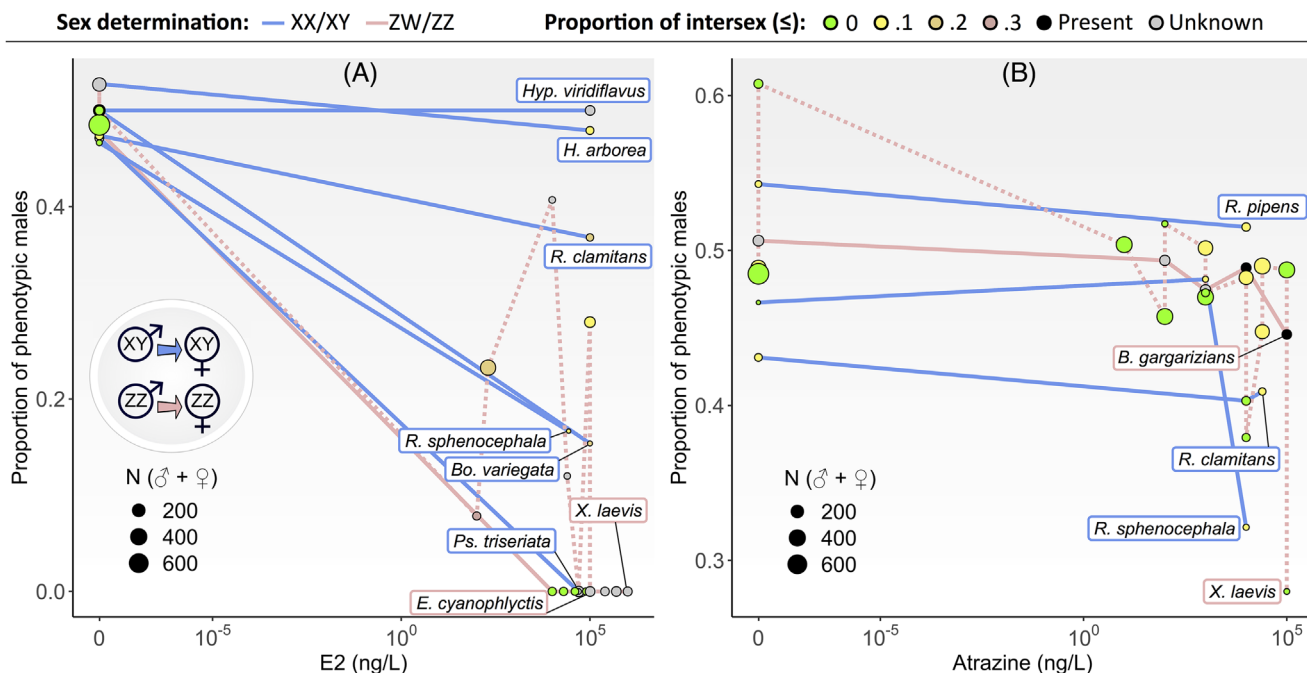


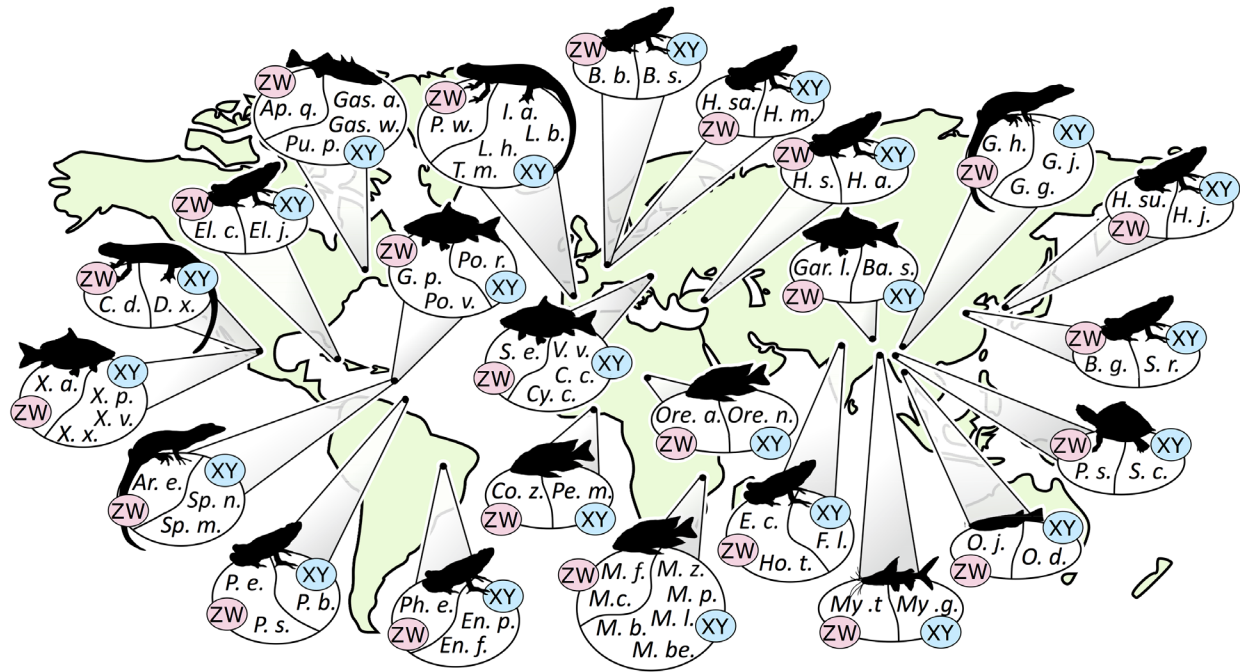
FIGURE 3 Phenotypic sex ratios reported from anurans exposed to different concentrations of E2 (A) or atrazine (B) as tadpoles. Within each panel, dot sizes are proportional to the number of animals with unambiguous sexual phenotype; dot colors indicate the proportion of intersex individuals among all animals examined for intersexuality. Supposed direction of sex reversal across both panels is denoted in the large white circle in Panel (A), where the symbols ♂ and ♀ stand for male and female phenotype, respectively. Displayed anuran genera are: *Bombina* (Bo.), *Bufo* (B.), *Euphyctis* (E.), *Hyperolius* (Hyp.), *Pseudacris* (Ps.), *Rana* (R.) and *Xenopus* (X.). For better visualization, data of *X. laevis* is connected with a dotted line. For data and references see Supplementary Table 1

shortly after metamorphosis in amphibians. The timing of dissection may significantly influence the results of sexing, because gonads in several amphibian and fish species undergo an ovary-like phase before differentiating into ovaries or testes, and the pace of this process also differs between species^[3,60,68,69] or even within the same species.^[70] Furthermore, the relative pace of gonadal and somatic development may vary between species and treatments^[71,72]; thus, treatment effects on somatic development (e.g., earlier metamorphosis at high temperatures) may lead to premature dissection and thereby sex assignment may be false^[12] or impossible (undifferentiated gonads: Supplementary Table 1). Apparently the same methodological issues led to earlier conclusions that sex reversal was only temporary in some amphibians.^[53,73] Therefore, for phenotypic sexing to be reliable, it should be performed at a sufficiently late age, which is usually well after metamorphosis in amphibians.^[71]

Because temperature can affect sexual development,^[7,16,57] experiments on chemically induced sex reversal should also pay attention to rearing temperatures. On the one hand, different species may adapt to different temperatures^[74]; thus, keeping the animals within their range of optimal temperatures is favored to prevent unexpected sex reversals or the above-mentioned methodological problems of sex-biased mortality and premature dissection. On the other hand, temperature may affect the solubility, uptake and degradation rate of the administered chemicals and ultimately their effects on sex.^[57,75,76] Therefore, rearing temperatures should be monitored, taken into account, and reported even when temperature effects are not the

focus of the experiment. Different species and even populations within species might differ in their sex-reversal propensity regardless of their sex-determination system. Local or species-specific adaptations in various traits may be developed to better survive and exploit conditions that vary across habitats, such as temperature^[74] or anthropogenic chemical pollution^[77]; similar adaptations might also increase or decrease the likelihood of sex reversal.^[41] Therefore, the source of the experimental animals, such as the climatic and land-use conditions of the collection sites or the specificities of the used breeding stocks (e.g., in *Xenopus*), should be clearly described in sex-reversal studies. When the experiments include both ZW/ZZ and XX/XY species, ideally these should be collected from the same sources or from similar habitats in order to improve the comparability of the two systems' response to specific sex-reversing conditions.

Also, earlier information may need revisiting and updating. For example, we recently found that sex ratio in the toad *Bufo bufo* (ZW/ZZ) was not affected by exposure to 30°C, a temperature high enough to cause up to 100% female-to-male sex reversal in the frog *Rana dalmatina* (XX/XY) in the same experiment.^[51] This contradicted the broadly cited study by Piquet,^[53] where 25°C produced male excess in *Bufo vulgaris* (a synonym for *Bufo bufo*). The contradiction can be resolved by realizing that Piquet captured her animals near Geneva, in a hybrid zone of *Bufo bufo* and *Bufo spinosus*,^[78] two species that were thought to be one at the time.^[79,80] Notably, *Bufo spinosus* features XX/XY system.^[81] Thus, it is likely that the study of Piquet^[53] is incorrectly cited in several reviews (e.g.,^[3,21,35]) as evidence for



Beloniformes

O. d. - *Oryzias dancena*
O. j. - *Oryzias javanicus*

Perciformes

Co. z. - *Coptodon zillii*
M. b. - *Metriaclima barlowi*
M. be. - *Metriaclima benetos*
M. c. - *Metriaclima callainos*
M. f. - *Metriaclima fainzilberii*
M. l. - *Metriaclima lombardoi*
M. p. - *Metriaclima phaeos*
M. z. - *Metriaclima zebra*
Ore. a. - *Oreochromis aureus*
Ore. n. - *Oreochromis niloticus*
Pe. m. - *Pelmatolapia mariae*

Siluriformes

My. t. - *Mystus tengara*
My. g. - *Mystus gulio*

Cypriniformes

Ba. s. - *Barilius shacra*
C. c. - *Carassius carassius*
Cy. c. - *Cyprinus carpio*
G. p. - *Gambusia puncticulata*
Gar. l. - *Garra lamta*
Po. r. - *Poecilia reticulata*
P. v. - *Poecilia velifera*
S. e. - *Scardinius erythrophthalmus*
V. v. - *Vimba vimba*
X. a. - *Xiphophorus alvarezii*
X. p. - *Xiphophorus pygmaeus*
X. v. - *Xiphophorus variatus*
X. x. - *Xiphophorus xiphidium*

Gasterosteiformes

A. q. - *Apeltes quadracus*
Gas. a. - *Gasterosteus aculeatus*
Gas. w. - *Gasterosteus wheatlandi*
Pu. p. - *Pungitius pungitius*

Caudata

C. d. - *Chiropoteritron dimidiatus*
D. x. - *Dendrotriton xoloccalcae*
I. a. - *Ichthyosaura alpestris*
L. b. - *Lissotriton boscai*
L. h. - *Lissotriton helveticus*
P. w. - *Pleurodeles waltl*
T. m. - *Triturus marmoratus*

Anura

B. b. - *Bufo bufo*
B. g. - *Bufo gargarizans*
B. s. - *Bufo spinosus*
El. c. - *Eleutherodactylus cuneatus*
El. j. - *Eleutherodactylus johnstonei*
En. f. - *Engystomops freibergeri*
En. p. - *Engystomops petersi*
E. f. - *Euphylyctis cyanophlyctis*
F. l. - *Fejervarya limnocharis*
Ho. t. - *Hoplobatrachus tigerinus*
H. a. - *Hyla arborea*
H. j. - *Hyla japonica*

H. m. - *Hyla meridionalis*

H. sa. - *Hyla sarda*

H. s. - *Hyla savignyi*

H. su. - *Hyla suweonensis*

Ph. e. - *Physalaemus ephippifer*

P. b. - *Pristimantis boconoensis*

P. e. - *Pristimantis euphonides*

P. s. - *Pristimantis shrevei*

S. r. - *Strauchbufo raddei*

Squamata

Ar. e. - *Aristelliger expectatus*

G. g. - *Gekko gekko*

G. h. - *Gekko hokouensis*

G. j. - *Gekko japonicus*

Sp. n. - *Sphaerodactylus nicholsi*

Sp. m. - *Sphaerodactylus macrolepis*

Testudines

P. s. - *Pangshura smithii*

S. c. - *Siebenrockiella crassicolis*

FIGURE 4 Examples of closely related male-heterogametic (XY) and female-heterogametic (ZW) species that occur in the same or relatively close geographical regions. Species in each oval belong to a single family, representing altogether nine orders of fish, amphibians and reptiles (denoted by different black icons). Data on sex determination was collected from [46,48,81,85–89], and data on taxonomy and indicative geographical distribution were obtained from [90,91]

temperature-induced sex reversal in *Bufo bufo*, and her findings potentially reflect female-to-male sex reversal induced in homogametic XX instead of heterogametic ZW individuals. With recent developments in molecular methods and resulting increase in the quantity and quality of data on taxonomy, sex chromosomes, and genetic sex markers, the time is now ripe for clarifying the role of heterogamety in sex-reversal susceptibility across ectothermic vertebrates.

CONCLUSIONS

Heterogamety is a fundamental aspect of organismal biology that, according to recent research, has far-ranging consequences on life histories and population dynamics, including sex ratios, sex-specific

aging rates, and life spans.^[82–84] Here we have highlighted that heterogamety may further influence the fate of ectothermic vertebrates by affecting their propensity to undergo environmental sex reversal. By considering “asymmetrical sex reversal,” a relaxed interpretation of Witschi’s rule, we can generate testable predictions regarding the differences in sex-reversal propensity between populations with different sex-chromosome systems induced by different environmental conditions. Empirical tests of these predictions are promising but so far scanty and difficult to integrate due to methodological heterogeneity behind the currently available results. Still, multiple findings suggest that in taxa like anurans and fish where high temperatures usually cause female-to-male sex reversal,^[1,3,9] climate change and urban heat islands may potentially pose greater risk to XX/XY compared to ZW/ZZ systems. By contrast, ZW/ZZ species may be more vulnerable to sev-

eral chemical pollutants that can induce male-to-female sex reversal. Furthermore, in species where temperature elevation induces male-to-female sex reversal (e.g., some reptiles^[2] and caudate amphibians^[16]), ZW/ZZ systems may be more threatened by heat waves. Therefore, more research on sex reversal is needed in order to assess the vulnerability of ectotherms to both climate change and environmental pollution. We hope that, by considering the methodological guidelines that we have provided in the present article, future studies will enable a systematic comparison of sex-reversal propensities in different sex-chromosome systems, and help to better understand the evolution of sex determination and its consequences in the Anthropocene.

SUPPORTING INFORMATION

Detailed searching methods are described in Supplement 1. Detailed references and the collected data on sex reversal, sex bias and sex determination are shown in Supplementary Table 1.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

All data and related references are included in the article/supplementary material.

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